

# Prostaglandin J<sub>2</sub> and Its Metabolites Promote Neurite Outgrowth Induced by Nerve Growth Factor in PC12 Cells

Takumi Satoh,\* Kyoji Furuta,† Masaaki Suzuki,† and Yasuyoshi Watanabe\*.1

\*Department of Neuroscience, Osaka Bioscience Institute, 6-2-4 Furuedai, Suita-shi, Osaka 565-0874, Japan; and †Department of Biomolecular Science, Faculty of Engineering, Gifu University, Gifu 501-1193, Japan

Received March 11, 1999

Although A- and J-type prostaglandins (PG's) arrest the cell cycle at the G1 phase in vitro and suppress tumor growth in vivo, their effects on neuronal cells have not so far been clarified. Here, we found promotion of neurite outgrowth as a novel biological function of PGJ's. In PC12h cells, PGJ's (PGJ<sub>2</sub>,  $\Delta^{12}$ -PGJ<sub>2</sub> and 15-deoxy-\(\Delta^{12,14}\)-PGJ2) promoted neurite outgrowth in the presence of nerve growth factor (NGF), whereas they themselves did not show such a promotion. The potency of promoting neurite outgrowth was PGJ<sub>2</sub> <  $\Delta^{12}$ -PGJ<sub>2</sub> < 15-deoxy- $\Delta^{12,14}$ -PGJ<sub>2</sub>. However, troglitazone, an activator of peroxisome proliferator-activated receptory (PPARy), and other PG's including PGA<sub>1</sub>, PGA<sub>2</sub> and PGD<sub>2</sub> did not promote neurite outgrowth. These results suggest that PGJ's promote neurite outgrowth independently of PPARy activation. © 1999

Key Words: prostaglandin J<sub>2</sub>; prostaglandin D<sub>2</sub>; peroxisome proliferator-activated receptor  $\gamma$ ; neurite outgrowth; PC12 cells; nerve growth factor.

Cyclopentenone PG's such as  $\Delta^{12}$ -PGJ $_2$  and PGA $_2$ reportedly inhibit cell proliferation and induce various biological activities including anti-viral activity and osteogenesis (1, 2). These PG's have no cell-surface receptors but are actively transported into cells and accumulate in nuclei (3-5). Although their nuclear receptor remains unclear, they induce a variety of genes such as p21  $^{\text{WAF1/CIP}}$  (6–8), p53 (9), *c-fos* (10), heme oxygenase-1 (11), HSP70 (12), IGF-1 (13), E-cadherin (14) and BIP/GRP78 (15). Recently, PGJ's (Fig. 1) were identified as natural ligands for PPARy and found to promote adipocyte differentiation (16, 17). The potency

<sup>1</sup> To whom correspondence should be addressed. Fax: +81-6-6872-0240. E-mail: watanabe@obi.or.jp.

Abbreviations used: DF medium, 1:1 mixture of Dulbecco's Modified Eagle medium and F12 medium; NGF, nerve growth factor; PG, prostaglandin; PPAR $\gamma$ , peroxisome proliferator-activated receptor  $\gamma$ .

of activating PPAR $\gamma$  is 15-deoxy- $\Delta^{12,14}$ -PGJ $_2 > \Delta^{12}$ -PGJ $_2$ > PGJ₂ > PGD₂. These results indicate that PGJ's are not only anti-tumor agents but also play important physiological roles since PGD<sub>2</sub>, a PGJ<sub>2</sub> precursor, is the most abundant PG in the CNS (18). PGD, is now wellknown as a sleep inducer in mammalian brain (19), but the neuronal effects of PGJ's are not clear. Thus we addressed the question as to what biological function PGJ's possess in neuronal cells. Since PC12 cells serve as a useful model system for the study of neurite outgrowth (20), we used this system to test the neuronal function of PGJ's.

## MATERIALS AND METHODS

Materials. NGF (2.5S) was purchased from Chemicon International. PGs (PGA<sub>1</sub>, PGA<sub>2</sub>, PGD<sub>2</sub>, PGJ<sub>2</sub>, Δ<sup>12</sup>-PGJ<sub>2</sub> and 15-deoxy-Δ<sup>12,14</sup>-PGJ<sub>2</sub>) (Fig. 1) were obtained from Cayman Chemical. Troglitazone was a generous gift from Sankyo Co. These compounds were diluted in ethanol. The final concentration of ethanol was 0.1% and this concentration did not affect neurite outgrowth or cell survival.

Cell cultures. PC12h cells, a subline of PC12 cells, were maintained in 75 cm<sup>2</sup> flasks (Costar) containing 1:1 mixture of Dulbecco's Modified Eagle medium and F12 medium supplemented with 5% (v/v) of heat-inactivated (56°C, 30 min) horse serum (GIBCO) and 5% (v/v) of precolostrum newborn calf serum (Mitsubishi Kasei) (5/5 DF).

For evaluation of neurite outgrowth, the cells were transferred to collagen-coated 24 well plates (Costar) at a density of 1x10<sup>4</sup> cells/cm<sup>2</sup> and incubated in 5/5 DF for 3 h. The medium was changed to serum-free DF, and the cells were incubated for 1 h, after which PG's were added. Thirty minutes later NGF (50 ng/ml) was added. The cells were then incubated for 24 h, and neurite (>10  $\mu$ m)-bearing cells per total cells (around 70-100) in the same area were counted.

## **RESULTS**

We examined the effects of PGJ's on neurite outgrowth induced by NGF in PC12h cells (Fig. 2). PC12h cells did not display any neurites when incubated in serum-free DF medium for 24 h (Fig. 2a). 15-deoxy-Δ<sup>12,14</sup>-PGJ<sub>2</sub> alone did not have any significant promoting effect (Fig. 2b). Incubation of PC12h cells with NGF



FIG. 1. Chemical structures of PG's used in the present study.

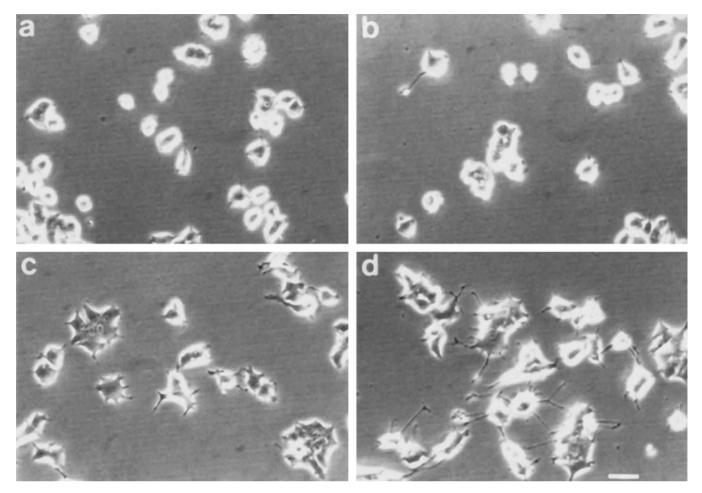
(50 ng/ml) for 24 hours induced neurite formation only slightly (Fig. 2c) since several days are required for neurite outgrowth in response to NGF. The presence of both 15-deoxy-Δ<sup>12,14</sup>-PGJ<sub>2</sub> and NGF potently promoted neurite outgrowth (Fig. 2d). We also examined the effects of PGJ's, PGA's, PGD<sub>2</sub> and troglitazone, an activator of PPARy (Fig. 3). Troglitazone, PGA<sub>1</sub>, PGA<sub>2</sub> and PGD<sub>2</sub> did not promote neurite outgrowth at 0.1-5.0  $\mu$ M, whereas PGJ<sub>2</sub>,  $\Delta^{12}$ -PGJ<sub>2</sub>, and 15-deoxy- $\Delta^{12,14}$ -PGJ<sub>2</sub> did promote the outgrowth. The potency of promoting neurite outgrowth was 15-deoxy- $\Delta^{12,14}$ -PGJ<sub>2</sub> >  $\Delta^{12}$ -PGJ<sub>2</sub> > PGJ<sub>2</sub> > PGD<sub>2</sub>. The promotion was observed within 48 hours, indicating that they only accelerated the speed of neurite outgrowth. Since higher concentrations (>0.5  $\mu$ M 15-deoxy- $\Delta^{12,14}$ -PGJ<sub>2</sub>; >2.0  $\mu$ M  $\Delta^{12}$ -PGJ<sub>2</sub> and PGJ<sub>2</sub>) induced cell damage, we could not count the number of neurite-bearing cells in cultures containing these high concentrations of PG's.

## **DISCUSSION**

The precursor of PGJ's, PGD<sub>2</sub>, is the most abundant cyclooxygenase product in the rodent brain (18). PGD<sub>2</sub>

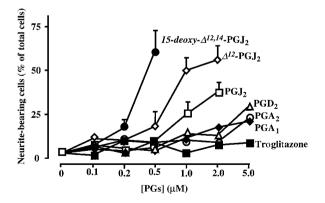
is sequentially metabolized to  $PGJ_2$ ,  $\Delta^{12}$ - $PGJ_2$ , and 15deoxy- $\Delta^{12,14}$ -PGJ<sub>2</sub> (2, 16, 17). PGJ's promoted neurite outgrowth induced by NGF (50 ng/ml) in PC12h cells (Fig. 2). The potency of promoting neurite outgrowth was 15-deoxy- $\Delta^{12,14}$ -PGJ<sub>2</sub>  $> \Delta^{12}$ -PGJ<sub>2</sub> >PGJ<sub>2</sub> >PGD<sub>2</sub> (Fig. 3), indicating a gain in biological potency as the catabolism of PGD2 proceeds. Since PGJ2 is easily converted to  $\Delta^{12}$ -PGJ<sub>2</sub>, the activity of PGJ<sub>2</sub> may be mediated by  $\Delta^{12}$ -PGJ<sub>2</sub>. If it is the case, the dienone structure of 15-deoxy- $\Delta^{12,14}$ -PGJ<sub>2</sub> and  $\Delta^{12}$ -PGJ<sub>2</sub> may be critical for promoting neurite outgrowth. The effective concentrations were 0.1–1.0  $\mu$ M (Fig. 3), which were about ten times lower than those required for growth arrest and induction of various genes (6-15). Although the function of PGJ's in the brain is not clear yet, our results suggest that promoting neurite outgrowth is an intrinsic biological activity of PGJ's in neurons.

Forman et al. (16) and Kliewer et al. (17) reported that PGD<sub>2</sub> metabolites bind and activate PPAR $\gamma$  and promote adipocyte differentiation, indicating that they are critical regulators of the generation of adipocyte *in vivo*. The potency of activating PPAR $\gamma$  is 15-deoxy-



**FIG. 2.** Promotion of neurite outgrowth of PC12h cells induced by NGF and 15-deoxy- $\Delta^{12.14}$ -PGJ $_2$ . PC12h cells were cultured in serum-free DF in the absence (a and b) or in the presence of NGF (50 ng/ml) (c and d) for 24 h. In (b) and (d), 15-deoxy- $\Delta^{12.14}$ -PGJ $_2$  (0.5  $\mu$ M) was added 30 min before NGF challenge. The bar represents 10  $\mu$ m.

 $\Delta^{12,14}$ -PGJ<sub>2</sub> >  $\Delta^{12}$ -PGJ<sub>2</sub> > PGJ<sub>2</sub>  $\gg$  PGD<sub>2</sub>. This potency order is the same as that in the promotion of neurite outgrowth in the present study, suggesting that they induce the activity through PPARy activation. However, the expression of PPARy is highly specific to adipotissue (21), suggesting that PC12 cells do not express it. In addition, troglitazone, an activator of PPARγ, did not promote neurite outgrowth up to 10  $\mu M$  (Fig. 3), indicating that PPAR $\gamma$  activation is not involved in the promotion of neurite outgrowth. It remains unclear yet as to what signaling pathway is required for the promotion of neurite outgrowth. PGJ's modulate expression of various genes, one of which may be essential for the promotion of neurite outgrowth in PC12 cells. One possible candidate is p21 WAF1/CIP1, because the overexpression of this gene reportedly accelerated neuronal differentiation in PC12 cells (22). Otherwise, PGJ's may activate MAP kinase activity, which is critical for neuronal differentiation in PC12h cells (23).



**FIG. 3.** Concentration-dependent promotion of neurite outgrowth in PC12h cells induced by PGJ's, PGA's and PGD<sub>2</sub>. PC12h cells were cultured in the serum-free DF in the presence of NGF (50 ng/ml) with various concentrations of PG's for 24 h. Neurite (>10 μm)-bearing cells per total cells (around 70–100) in the same area were counted. The values are means  $\pm$  S.D. (n = 4). Closed circles, 15-deoxy- $\Delta^{12.14}$ -PGJ<sub>2</sub>; open diamonds,  $\Delta^{12.14}$ -PGJ<sub>2</sub>; open squares, PGJ<sub>2</sub>; open triangles, PGD<sub>2</sub>; open circles, PGA<sub>2</sub>; closed squares, troglitazone.

In conclusion, we found that PGJ's promoted neurite outgrowth induced by NGF in PC12 cells independently of PPAR $\gamma$  activation. This result suggests the possibility that PGJ's are critical regulators of neurite outgrowth or regeneration in the nervous system.

#### **ACKNOWLEDGMENTS**

This work was supported in part by the Research for the Future Program (RFTF) JSPS-RFTF 98L00201 from the Japan Society for the Promotion of Science (JSPS) (Y.W.) and a Grant-in-Aid for Scientific Research on Priority Area No. 09273102 from the Ministry of Education, Science, Sports and Culture (M.S.). The authors thank Dr. Koichi Kojima of Sankyo Co. for the generous gift of troglitazone and Dr. Larry D. Frye for editorial help with the manuscript.

#### REFERENCES

- Honn, K. V., Bockman, R. S., and Marnett, L. J. (1981) Prostaglandins 21, 833–864.
- 2. Fukushima, M. (1992) Prostaglandins Leukotrienes and Essential Fatty Acids 47, 1–12.
- 3. Narumiya, S., and Fukushima, M. (1986) *J. Pharmacol. Exp. Ther.* **238**, 500–505.
- Narumiya, S., Ohno, K., Fujiwara, M., and Fukushima, M. (1986) J. Pharmacol. Exp. Ther. 238, 506-511.
- Narumiya, S., Ohno, K., Fukushima, M., and Fujiwara, M. (1987) J. Pharmacol. Exp. Ther. 242, 306–311.
- Gorospe, M., Wang, X., Guyton, K. Z., and Holbrook, N. J. (1996)
  Mol. Cell. Biol. 16, 6654–6660.
- Tanikawa, M., Yamada, K., Tominaga, K., Morisaki, H., Kaneko, Y., Ikeda, K., Suzuki, M., Kiho, T., Tomokiyo, K., Furuta, K., Noyori, R., and Nakanishi, M. (1998) J. Biol. Chem. 273, 18522– 18527.

- 8. Ishikawa, T., Akimaru, K., Nakanishi, M., Tomokiyo, K., Furuta, K., Suzuki, M., and Noyori, R. (1998) *Biochem. J.* **336**, 569–576.
- 9. Lee, J. H., Kim, H. S., Jeong, S. Y., and Kim, I. K. (1995) *FEBS Lett.* **368**, 348–352.
- 10. Higashiyama, K., Niiya, K., Ozawa, T., Hayakawa, Y., Fujimaki, M., and Sakuragawa, N. (1996) *Prostaglandins* **52**, 143–156.
- Negishi, M., Odani, N., Koizumi, T., Takahashi, S., and Ichikawa, A. (1995) FEBS Lett. 372, 279–282.
- Holbrook, N. J., Carlson, S. G., Choi, A. M. K., and Fargnoli, J. (1992) Mol. Cell. Biol. 12, 1528-1538.
- Bui, T., Kuo C., Rotwein, P., and Straus, D. S. (1996) Endocrinology 138, 985–993.
- Ikai, K., Yamamoto, M., Matsuyoshi, N., and Fukushima, M. (1995) Prostaglandins Leukotrienes and Essential Fatty Acids 52, 303–307.
- Odani, N., Negishi, M., Takahashi, S., Kitano, Y., Kozutsumi, Y., and Ichikawa, A. (1996) *J. Biol. Chem.* 271, 16609–16613.
- Forman, B. M., Tontonoz, P., Chen, J., Brun, R. P., Spiegelman, B. M., and Evans, R. (1995) Cell 83, 803–812.
- Kliewer, S. A., Lenhard, J. M., Willson, T. M., Patel, I., Morris,
  D. C., and Lehmann, J. M. (1995) *Cell* 83, 813–819.
- Gaudet, R. J., Alam, I., and Levine, L. (1983) J. Neurochem. 35, 653–658.
- 19. Hayaishi, O. (1997) *in* Sleep–Wake Disorders (Meier-Ewert, M., and Okawa, M., Eds.), pp. 1–10, Plenum Press, New York.
- Greene, L. A., and Tischler, A. S. (1976) Proc. Natl. Acad. Sci. USA 73, 2424–2428.
- Chawla, A., Schwarz E. J., Dimaculungun, D. D., and Lazer, M. A. (1994) Endocrinology 135, 798–800.
- 22. Erhardt, J. A., and Pittman, R. N. (1998) Oncogene 16, 443-451
- Segal, R., and Greenberg, M. E. (1996) Annu. Rev. Neurosci. 19, 463–489.